

MICROCOPY

CHART



NRL Memorandum Report 5744

# Advantages of the Gas Exchange Approach to Microbiological Studies

P. J. HANNAN AND D. S. JONES

Combustion and Fuels Branch Chemistry Division

April 3, 1986





NAVAL RESEARCH LABORATORY Washington, D.C.

OTIC FILE COPY

AD-A166 887

Approved for public release; distribution unlimited. 86

073

_			
SECTION TV	CLASSISIO	TION OF	THIS PAGE

AD-A166 887

SECURITY CLAS	ISICIONI UN UI	100	AUE	REPORT DOCUM	AENTATION!	DAGE	4		
				REPORT DOCOM					
UNCLASSI		FICATIO	; N		16 RESTRICTIVE	MARKINGS			
2a. SECURITY	CLASSIFICATION	HTUA P	ORITY		3 DISTRIBUTION	/AVAILABILITY OF	RSPO	RT	
26. DECLASSIF	ICATION / DOW	NGRAD	ING SCHEDU	.E	Approved for	public release;	distr	ibution u	nlimited.
4. PERFORMIN	G ORGANIZAT	ON REP	ORT NUMBER	R(S)	5 MONITORING	ORGANIZATION RE	PORT	NUMBER(S)	
NRL Memo	orandum Re	port 5	7-14						
6a. NAME OF	PERFORMING	ORGANI	ZATION	6b. OFFICE SYMBOL	7a NAME OF ME	ONITORING ORGAN	IZATI	ON	
Naval Resea	arch Labora	tory		(If applicable)					
6c. ADDRESS (	City, State, and	ZIP Co	de)		75. ADDRESS (Cit	y, State, and ZIP C	ode)		
Washington	, DC 2037	5-5000	)						
8a. NAME OF ORGANIZA		NSORIN	G	8b. OFFICE SYMBOL (If applicable)	9. PROCUREMEN	T INSTRUMENT IDE	NTIFIC	ATION NUI	MBER
	cal R&D Co	mman	d	(ii) applicable)	[				
8c. ADDRESS (				<u> </u>	10 SOURCE OF	UNDING NUMBERS			
	-				PROGRAM	PROJECT	TASK		WORK UNIT
Bethesda, N	40 20014				ELEMENT NO.	NO	NO.		ACCESSION NO.
			<del></del>	· · · · · · · · · · · · · · · · · · ·	(See page ii)	<u> </u>			L
11 TITLE (Incli	-			ach to Microbiologie	al Studios				
Advantages	of the Gas	EXCIIA	nge Appros	ich to Microbiologie	ai Studies				
12. PERSONAL		ъ с							
Hannan, P.		s, D. S.							
13a. TYPE OF Interim	REPORT		13b. TIME CO	983TO1985	14. DATE OF REPO	RT (Year, Month, D	ay)	15. PAGE 0 45	COUNT
16 SUPPLEME	NTARY NOTAT	ION				_ <del>`</del>			
17	COSATI	CODES		18. SUBJECT TERMS Y	ontinue on revers	e if necessary and	ident	ify by block	number)
FIELD	GROUP	SUB	3-GROUP	Gas exchange	Chlorella,	O2 production	n į		
				Algae	Toxicity	CÖ2 uptake,			
				L					·
			<del>-</del>	and identify by block n					
Studies o	f the effects	of var	rious chem	ical or physical stim	uli on the grow	th rates of micr	oorga	anisms ger	aerally involve
some measu	re of bioma	ss. In	the case of	algae, one might m	easure the cell r	rumber, the chi	orop	nyll conce	entration, the
Huorescence	e, or the we	weigh	nt of the cu	ilture as a function d	of time. Each s	uch measureme	nt re	quires a sa	impling of the
culture which	ch could be	a distu	irbing facto	or in the system bein	ig measured. A	nother disadvar	itage	of these t	raditional
furthermore	its is that the	ne tim	es requirea	for significant chan	ge to take place	e in the culture	migh	t be hours	s or days:
				calculated from the vas constant.	data must be b	ased on the assu	ımpt:	ion that d	uring the time
								<b>H</b>	}
tion of an ai	ir etroam na	anotr wina H	er approaci brough tho	n to the problem. It system (the <sup>⊊</sup> gas ex	consists simply	y of monitoring	the (	Og or CO	2 concentra-
1) there is no	n sercam par o need to tak	osung u to sam	nles of the	culture, 2) each mea	cuange metno	at, and it has so	vera	. distinct a	advantages:
ance of the	culture at th	at ver	v moment.	and 3) transitory el	nanges in growt	h ratos aro roadi	ite de	u muncani	S the perform-
given of stud	dies made p	revious	sly with thi	s method at NRL.	Also the possible	le value of these	met	hods in a	study of the
corrosion su	sceptibility	of allo	ys is descr	ibed. (** 🚉 📉 🗥	٠ <u>٠</u> ٠				
	TION / AVAILAB		F ABSTRACT  SAME AS F	RPT DTIC USERS	21. ABSTRACT SE UNCLAS	CURITY CLASSIFICA	ATION		
22a. NAME O				TO THE OSERS		(Include Area Code)	1 220	OFFICE SY	MBOL
Patrick J. H				بيانا المام والمرابع والمسيون	(202) 76		1	ode 6180	

DD FORM 1473, 84 MAR

83 APR edition may be used until exhausted All other editions are obsolete.

SECURITY CLASSIFICATION OF THIS PAGE

### SECURITY CLASSIFICATION OF THIS PAGE

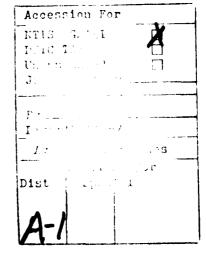
CONTRACTOR OF THE PROPERTY OF

10. SOURCE OF FUNDING	NUMBERS		
PROGRAM ELEMENT NO.	PROJECT NO.	TASK NO.	WORK UNIT ACCESSION NO.
61153N 62000N	SP 89422	CO8-32	DN480-754

### CONTENTS

I.	INTRODUCTION	]
II.	MATERIALS AND METHODS	2
		2
	Culture Devices	2
	All flow measurements and Gas Analyses	•
III.	RESULTS	
	Effect of Light Intensity on CO2 Uptake	_
	and O <sub>2</sub> Production	5
	Effect of CO <sub>2</sub> Input Rate	ŧ
	Effect of Additional Urea on Gas	_
	Exchange Rates	ζ
	Culture Medium	,
	Effect of Pressure on O <sub>2</sub> Production	-
	Toxicity of Metal Alloys to Chlorella	8
	Reliability of Algal Cultures	9
	Limitations of Gas Exchange Measurements with	
	Water of High Salinity	L (
IV.	DISCUSSION	13
	Reproducibility	1:
	Additional Factors to Be Considered	
	Alloy Toxicity	LS
٧.	REFERENCES	٦.





# ADVANTAGES OF THE GAS EXCHANGE APPROACH TO MICROBIOLOGICAL STUDIES

### I. INTRODUCTION

The purpose of this report is to point out a unique aspect of a particular biological measurement and relate it to possible future studies at the Naval Research Laboratory. The methodology is relatively simple but it has not been used to the extent that its potential warrants.

The technique consists in monitoring, on a continuing basis, the CO<sub>2</sub> (or O<sub>2</sub>) content of an air stream following its passage through a culture of microorganisms. If the air flow rate and the temperature of the liquid are constant, then a comparison of the input and exit gases provides an instantaneous measure of the growth rate of the culture. Transient changes in growth rates, resulting from the imposition of various stimuli, can be reflected in changes in gas composition; by choosing an organism which is particularly susceptible to a certain stimulus, the technique can have extreme sensitivity. This report will describe the research done previously at NRL which illustrates the rapid sensing of changes in growth rates based on gas exchange. Algal cultures have been the subject of these studies and in certain instances both CO2 and O2 measurements have been made; the same techniques could be used for studies of other microorganisms provided they have a sufficiently fast growth rate. An inherent advantage of CO2 measurements is that they can be made against a background of atmospheric CO2 concentrations which are several orders of magnitude less than O2 concentrations.

Traditional measurements of microorganism growth rates can be based on actual counts of cells, or analyses for adenosine triphosphate (ATP), chlorophyll, or other products, at various intervals of time. The time between measurements must be sufficient to allow significant changes to take place in the parameter being measured; small differences between large numbers are often encountered. By contrast, each gas exchange measurement is a rate measurement and the problem of a large background can be minimized. To illustrate the point, consider a hypothetical algal culture being aerated at a constant rate and diluted with fresh culture medium at a constant rate. system comes to equilibrium with a fixed cell density because the cells being washed out are replaced quantitatively with new cells, and the culture is producing O2 at a constant rate. Now, suppose the aeration and dilution rates are constant but the

Manuscript approved November 20, 1985.

growth rate is reduced 10% (perhaps by a lower light intensity). Figure 1 depicts the gradual change in cell concentration which does not come to a new equilibrium until 10 hours have elapsed, whereas the new gas exchange rate is stabilized in a matter of minutes. The time required for the new gaseous equilibrium depends partially on the flow rate. For CO<sub>2</sub> equilibria the salinity is an important factor, also, which is covered later in this report. The essential point is that the gas exchange approach tells, in a matter of minutes, what would require hours if biomass were being measured. An assumption regarding comparability between O<sub>2</sub> production and biomass production is made for the purpose of the illustration in Figure 1; the authors are aware of the inaccuracy but prefer to make the case in the simplest form at this point.

The gas exchange approach has been used extensively at the Naval Research Laboratory in studies with the freshwater alga, Chlorella pyrenoidosa sorokiniana. In the paragraphs following, data will be presented to show the reliability of such measurements and the speed with which they reflect changes in culture conditions such as light intensity, CO2 input rate, pressure, or the presence of toxic alloys. Interest in the gas exchange phenomenon began here with studies concerning the feasibility of providing oxygen for nuclear submarines by the use of mass cultures of algae. Many types of culture apparatus were used, each designed for a specific purpose. The final report on these studies stressed the reliability of the system but pointed out its impracticality because of the exorbitant electrical energy requirement (Hannan, Shuler, and Patouillet, 1963).

### II. MATERIALS AND METHODS

### Organism

Chlorella pyrenoidosa 7-11-05 (later given the sub-species name of sorokiniana) was isolated initially from a Texas swamp, and cultures were obtained from Dr. Constantine Sorokin while he was on the faculty of the University of Maryland. Its high growth rate, with a doubling time of only two hours under ideal conditions, made it particularly attractive for the purpose at hand (Sorokin and Myers, 1954). The organism is roughly circular with a diameter of approximately 5 microns, and has a temperature optimum of 37-39°C. Sterile conditions were not maintained, and tap water was used in the preparation of the medium (Table 1).

### Culture Devices

A. Six-lamp unit. Many experiments were performed with the large culture apparatus shown in Figure 2. It is described in detail elsewhere (Hannan and Patouillet, 1963 A), but the essential features were temperature control within 2°C, high

light intensities provided by six GE Quartzline lamps oriented vertically in the six liters of culture medium, rapid stirring, controlled rates of dilution with fresh culture medium, close control over the concentration of CO2 in the input air, and the rate of air input. Air was bubbled through the culture at rates up to 4 liters/min and allowed to escape through ports in the top of the unit. Sampling of the effluent air was by means of a small pump connected to a narrow-bore nylon tube which extended into the air space above the algal culture; the output of the pump (400 cc/min) was only a fraction of the air input rate to the system, therefore the effluent from the pump consisted solely of air that had passed through the culture. This six-lamp unit was used in the experiments described subsequently concerning the effects of light intensity, CO2 input rate, urea, and the 'Matthern' medium.\* It was used as the source of cells for experiments concerning the effects of pressure, and the toxicity of alloys, each of which required a different type of culture apparatus.

- Pressure Unit. This unit (Figure 3) was made from glass pipe sealed with rubber O-rings into blocks of lucite, and contained two annular spaces around a vertically oriented Quartzline lamp. Cooling water flowed through the annulus nearest the light, and the algal culture was pumped through the outer annulus by a gear pump. A constant pressure was maintained within the culture chamber by manipulating stainless steel needle valves which controlled the input and exits of the air/CO<sub>2</sub> mixtures used. The CO<sub>2</sub> input rates (i.e., the product of the air flow rate times the CO2 concentration) could be varied by changing the flow rate of a given air composition, or by changing to a different CO2/air mixture. Pressure cylinders of particular CO2/air mixtures were used in this study to assure constant CO2 concentrations throughout an experiment. In these experiments, there was no dilution of the cultures with fresh medium. A complete description of the apparatus and results can be found elsewhere (Hannan, 1964).
- C. Toxicity Unit. Various experiments were performed with this unit but the emphasis here will be placed on the experiments concerning the potential toxicities of various metal alloys. The unit used in this study (Figure 4) had two annular spaces surrounding a vertical Quartzline lamp, the inner containing cooling water and the outer containing the algal culture which was circulated by a gear pump. The outer shell of the unit was made of #304 stainless steel which, in turn, was surrounded by an annular space through which cooling water was circulated on demand from a thermoregulator extending into the culture. At the top of the annular space for the culture was an

<sup>\*</sup>See page 6

O-ring seal through which a 3.18 mm (1/8) inch diameter rod could be extended, the rod being representative of a particular alloy to be studied. In the initial phase of an experiment, a 350 ml portion of a fresh algal culture taken from the six-lamp unit was placed in the apparatus and its  $0_2$  production measured until a steady state had been achieved. Then the O-ring seal in the top of the unit was loosened slightly to permit lowering the rod into the culture, and  $0_2$  production measurements were continued. The length of rod in the culture was 20.3 cm (8). The results are described in the report by Hannan and Patouillet (1967). There was no dilution of the culture with fresh medium during these experiments.

### Airflow Rate Measurements and Gas Analyses

Flowmeters containing either a Pyrex or steel ball were used, depending on the flow rate being measured; the meters used were products of either Ace Glass or the Fischer and Porter Co. Air leaving the culture chamber was first passed through a bubbler\* and then a Drierite (anhydrous CaSO<sub>4</sub>) column to remove water vapor, before passing through the flowmeter.

The  $O_2$  analyzer was a Model F-3 (Beckman Instruments, CA) with two scales, 0-25% and 20-25%, the latter being the one generally used because estimates of  $O_2$  concentrations could be made to 0.01%. The  $CO_2$  analyzer was the product of Mine Safety Appliances (Pittsburgh, PA), with a single 0-2% range which was later modified to extend to 4%. Zero settings of each instrument were made with  $N_2$  passing through; the  $O_2$  analyzer was spanned with air (assumed to be 20.95%  $O_2$ ) and the  $CO_2$  analyzer was calibrated with standard gases whose  $CO_2$  content had been determined earlier with a modified Orsat apparatus. With this particular apparatus,  $CO_2$  analyses were reproducible within 0.02%.

In order for meaningful  $\mathrm{CO}_2$  uptake measurements to be made, it was necessary to control the input  $\mathrm{CO}_2$  concentration closely. Because the flow rates of air through the six-lamp unit were so high, it was not practical to use pre-mixed air from cylinders. Instead, a manifold was constructed to connect four  $\mathrm{CO}_2$  cylinders to a mixing system made at NRL (Bultman and Hannan, 1967) in which compressed air from the laboratory was mixed with  $\mathrm{CO}_2$  at a controlled rate by means of a needle valve on the  $\mathrm{CO}_2$  inlet. With this equipment, it was possible to maintain a  $\mathrm{CO}_2$  concentration within several hundredths of a percent for weeks at a time.

<sup>\*</sup>A small suction flask fitted with a rubber stopper holding a glass T; the lower leg of the T extended into a dilute acid solution. Air flowing through the exit of the T was controlled by a screw clamp to give the desired flow rate into the analyzer.

A particular characteristic of this unit was that the concentration of  ${\rm CO}_2$  was not affected by the rate at which the air was tapped off the mixing system.

For experiments conducted with the pressure unit or the toxicity unit, the  $CO_2/air$  mixtures were taken from cylinders containing known concentrations of  $CO_2$ .

The time required for the gas analyzers to equilibrate with an air stream depended on the flow rate of air, the volume of connector tubing and desiccant in the train of equipment leading to the analyzers, and the type of algal culture being used; the complication attendant with CO<sub>2</sub> uptake measurements concerned only high salinity, which is discussed at length later in this report. For the experiments with the fresh water alga, at a flow rate of 100 cc air/min, about eight minutes was required for the meters to reflect accurately the composition of the air; with the six-lamp unit, the higher flow rates of air required even less time for equilibrium.

### III. RESULTS

# Effect of Light Intensity on $CO_2$ Uptake and $O_2$ Production

Light intensity within the six-lamp unit could be changed quickly since one large variac controlled the voltage on the lamps. Figure 5 represents the change in CO2 uptake and O2 production with a change in voltage; with 200 volts on the lamps the O2 production was 3,400 cc/hr but when the voltage was dropped to 175 the  $O_2$  production became approximately 2,700. The new equilibrium value of the  $\mathrm{O}_2$  production was apparent in only two minutes, with a little more time required for the CO2 absorption to reach a new steady state. Several factors played roles in the times required for new equilibria to be established: 1) The high air flow rate through the culture was effective in stripping the dissolved  $O_2$  and  $CO_2$  from the culture and, 2) Equilibrium between gaseous  $CO_2$  and the liquid culture probably required more time than that of the O2 because of the salinity of the culture. This factor is explored in more detail later in this report. When the voltage on the lamps was restored to 200 the original  $O_2$  and  $CO_2$  values returned, once again with a slight delay in the case of the CO2.

It is possible, of course, to demonstrate a rapid change in O2 production with change in light intensity with the use of a polarograph. With a heavy deposit of algal cells placed directly on a polarographic electrode, changes in oxygen tension can be monitored with time but such a system does not provide the flexibility of studies which is possible with the gas exchange approach.

### Effect of CO2 Input Rate

It had been shown earlier that the rate of O2 production depended on the CO2 input rate, i.e., the product of the CO2 concentration times the flow rate. This can be detected almost immediately by the gas exchange approach, as shown by the plot in Figure 6. With a 3.8% CO2 concentration in an air flow at 2,200 cc/min, the  $CO_2$  input rate was 5,020 cc/hr and the  $O_2$ production rate was 3,380 cc/hr. By reducing the air flow rate to 1,743 cc/min the  $CO_2$  input rate became 3,975 and the  $O_2$ production dropped to 3,200. When the flow rate was increased again, the 02 production rose above the expected value and stayed there for approximately a half-hour before resuming the normal value for those conditions. This was probably a consequence of the photosynthetic potential developed in the cells when they were in a CO2-deficient status, resulting in a temporarily enhanced productivity when more  $CO_2$  became available. This transient condition would not likely be detectable by monitoring cell counts or chlorophyll content.

### Effect of Additional Urea on Gas Exchange Rates

Included among the attempts to maximize  $O_2$  production was an experiment to determine the effect of an increased nitrogen concentration in the culture medium. The six-lamp culture unit had been at equilibrium, while being diluted with fresh culture medium at 15%/hr, when solid urea was added in an amount calculated to double the concentration in the culture medium. Because of the fast dissolution of urea in this rapidly stirred suspension, the change in urea concentration was probably almost immediate.

The effect of this urea addition on gas exchange is illustrated in Figure 7. The gyrations in  $CC_2$  uptake and  $CC_2$  production are not understood but the major point to be stressed is that changes in packed cell volume (which were included in this study) were minor compared to the large changes in gas exchange. Within a four-hour period, the  $CC_2$  absorption had decreased by 7.9% and then increased by 3.9% during which time the packed cell volume had shown a slight steady increase. The last reading of the day indicated a marked decline in both  $CC_2$  uptake and  $CC_2$  production but by the next morning the cells had recovered and their production was normal. Dilution with fresh medium had continued overnight so that there was a dual effect on the system, i.e., a gradual washout of the excess urea and probably an acclimation, with time, of the cells to their changed environment.

### Adaptation to a More Concentrated Culture Medium

In another experiment to maximize  $O_2$  production by the Chlorella culture, a concentrated culture medium (devised by Dr. Robert Matthern at the Quartermaster Laboratories in

Chicago) was substituted for the normal medium. Within less than four hours, when the dilution rate was 15% hr, the  $\rm O_2$  production had decreased by one-third and within 20 hours it had dropped by almost one-half (Figure 8) at which point the dilution was stopped lest all the cells be washed out. Ultimately, the dilution was resumed but only at 3.3%/hr, then at 6.5% and finally 13% on the fourth day of the experiment. Changes in the gas exchange characteristics of the culture had provided an early warning of its potential demise in a manner that could not have been done with the cell-counting method. It was interesting that the culture finally achieved the normal gas exchange values with this concentrated medium and, also, that it maintained them when it was switched back to the regularly used medium.

### Effect of Pressure on O2 Production

A factor to be considered in an algal culture unit for submarine use would be the relatively low ambient concentrations of  $\text{CO}_2$  compared to those used in our experiments. The premise was that pressurization would increase  $\text{CO}_2$  solubility and, therefore, enhance  $\text{O}_2$  production. The equipment has already been described and is pictured in Figure 3.  $\text{CO}_2/\text{air}$  mixtures were supplied from cylinders having fixed  $\text{CO}_2$  concentrations and for each 10 psi increment studied, there were  $\text{CO}_2$  input rates of 30-, 60-, and 100% saturation at the light intensity used. A given culture was used for only one pressure study lest there be some residual effect of pressure, so that for each pressure condition there were three different batches of cells used.

The results are summarized in Figure 9. The  $O_2$  production, at time zero, represented the steady state value for the culture at atmospheric pressure with that particular  $CO_2$  input rate. With the imposition of pressure the  $O_2$  production generally changed and, except for the highest  $CO_2$  input rates used, there tended to be a temporary increase in  $O_2$  production followed by a decline: the higher the pressure imposed, the more drastic the decline in  $O_2$  production. Experiments with cellfree solutions showed that within 15 minutes a steady state had been established between the liquid and the  $O_2$  and  $CO_2$  in the air stream, therefore the  $O_2$  production and  $CO_2$  absorption values shown after that time were the result of biological action of the algal cells and not merely attributable to physical effects.

There was a possibility that this reduced  $O_2$  production was related to an increase in  $O_2$  tension in the liquid because the literature contains references to inhibitions obtained with algal growths subjected to high  $O_2$  concentrations. To test this hypothesis, a special mixture of  $CO_2$  (2.49%) and  $O_2$  (6.84%) in  $N_2$  was provided at a single  $CO_2$  input rate of 1,716 cc/hr, and at 10 to 50 psi. A plot of  $O_2$  production vs.

time under pressure was similar to that when  ${\rm air/CO_2}$  mixtures were used, indicating the oxygen tension was not a significant factor in the inhibitions described.

The principal point to be made is that the gas exchange rates changed quickly under pressure. It would not be possible to determine the magnitude of these transient changes by monitoring cell counts or other measures of biomass because there would need to be a constant productivity over a period of several hours to establish a definite relationship between gas exchange and biomass production.

### Toxicity of Metal Alloys to Chlorella

It is known that certain alloys are toxic, based on the comparative growth rates of algal cultures in the presence of these alloys (Dyer and Richardson, 1962). To build a large algal gas exchanger for submarine use would require that non-toxic structural materials be used, and the intent of this study was to explore further the range of options available; instead of the traditional approach used in the Dyer and Richardson study, the procedure was to introduce test alloys into a growing algal culture and determine their effect or the culture by monitoring gas exchange.

Each experiment consisted in establishing the O2 production of a batch culture of Chlorella, and then inserting into it a 3.8 mm diameter rod while continuing to monitor the O2 production. Figure 10 represents the data obtained with a rod made of #304 stainless steel; within 15 minutes after the insertion of the rod the productivity dropped to a lower plateau and remained This curve typifies the results obtained with a variety of alloys. Complete details of this study can be found elsewhere (Hannan and Patouillet, 1967) but the principal finding was that the surface properties of an alloy, and not its bulk composition, determine whether it will exert a toxic action on a culture. Many experiments were performed with the rod that was the subject of Figure 10, and eventually its toxicity was lost which would indicate that some leachable component had finally disappeared entirely from the surface. It was conceivable that this component(s) was being released from a grain boundary; to test this hypothesis, the specimen was stretched 1% of its length in a tensile test machine and then assayed again with a Chlorella culture. The toxicity of the rod was restored, the presumption being that a new grain boundary had been exposed from which toxicants could be released.

The most compelling evidence that the bulk composition was not the determinant in these toxicity studies was obtained with a 182.9 cm length of #316 stainless steel which was cut into seven 25.4 cm specimens tested. Four showed evidence of toxicity (ranging from 8 to 15% inhibition) while three had essentially no effect. Also pertinent to this theme of surface

composition as the determinant in activity were the results obtained with a brass rod which was assayed on four occasions after being buffed. The results ranged from a slight enhancement in productivity to 24% inhibition.

No provisions were made in this culture unit for continuous dilution with fresh medium, so it was not possible to determine whether the inhibitory effects might be of short or long duration. Beyond a period of about four hours, there would be the possibility that a reduction of  $O_2$  production would be the result of a nutrient deficiency which could be misinterpreted as a toxic effect.

### Reliability of Algal Cultures

In the preceding paragraphs, stress was placed on the speed with which changes in gas exchange rates can be detected following the addition of a stimulus to the cultures. The credibility of such data depends on the reliability of the method and examples will be shown here of a high degree of reproducibility obtained with different types of algal cultures used in different ways; the first concerns data on O2 production by the fresh water Chlorella, and the second relates to growth rate studies (chlorophyll production) made with the marine diatom, Phaeodactylum tricornutum.

The reliability of the six-lamp unit was determined in a series of measurements made over a five-day period in which the culture was operated at a fixed CO2 input rate and a fixed dilution rate, with eight measurements of O2 production and CO2 uptake made each day. For sixteen hours out of each day, there was no one in attendance. For the gas exchange measurements, it was necessary to calibrate the  $0_2$  and  $CO_2$  analyzers and connect the appropriate inlet and outlet sources to them at the proper times. The results are included in the report by Hannan and Patouillet (1963) and summarized in Table 2. high standard deviation in O2 production on the first day indicated the gradual acclimation of the culture to the conditions established, since the culture had been in the refrigerator during the weekend prior to the test. By the 5th day of the test, however, with an average O2 production rate of 3,638 cc hr<sup>-1</sup> the standard deviation was only 24, amounting to an error of less than 1%. This discrepancy could be caused by an error of only 0.01% in reading the  $O_2$  analyzer, and it was calibrated only in tenths.

The second example of reliability concerned a study of growth rates obtained with <u>Phaeodactylum tricornutum</u>, based on fluorescence measurements made after 20-and 44-hours of incubation. In these experiments, a standard toxicant was used, i.e., 3.3 parts per billion Hg (as HgCl<sub>2</sub>). Over a period of several months there were many experiments performed with a large batch of aged synthetic seawater, all performed with batch

cultures used in a definite regimen. The average inhibition of growth was 35% by the Hg, with a standard deviation of 4.3. This is considered an exceptionally low standard deviation for results with batch cultures used at different times. The sensitivity of the procedure is dramatized also when results are compared with a parallel study in which another batch of synthetic seawater was the base; here the inhibition with Hg was only 23%. While distilled water and C.P. chemicals were used for the preparation of both batches of seawater, there obviously was a difference in the results obtained. These experiments are described in detail in the report by Hannan and Patouillet (1979).

### <u>Limitations of Gas Exchange Measurements</u> with Water of High Salinity

The equilibration of  $\mathrm{CO}_2$  with water is rather rapid when the salinity is low, such as with the culture medium for  $\underline{\mathrm{Chlorella}}$ . With high salinities, however, such as with seawater, the equilibration rate is so slow that gas exchange measurements are not possible. The question then arises as to the maximum salinity with which meaningful  $\mathrm{CO}_2$  exchange measurements can be made.

To determine this, an apparatus was constructed which consisted of a long glass tube filled with water through which a known concentration of CO2 in air was bubbled until the water was saturated, i.e., the  $\bar{\text{c}}$ omposition of the effluent air was the same as the input. Then air of a different  $CO_2$  concentration was introduced and the time required for equilibration was In these experiments, several factors were kept constant; the air flow rate was 10 cc/min, the bubble diameter was 5 mm, and the average temperature was 22.44°C with a maximum variation of 0.06°C. The CO<sub>2</sub> concentrations used were 0- and 346 ppm. Four salinities were used, viz. 25-, 50-, 75-, and 100-% seawater which was prepared from distilled water and C.P. chemicals according to the formula of Lyman and Fleming (1940). An additional .144 gram NaHCO3/liter was added to Lyman and Fleming's formula for a purpose apart from the present The salinity of the 100% seawater was 32.18 parts discussion. per thousand

The 346 ppm  $\rm CO_2$  concentration used in these experiments was orders of magnitude less than those in the studies with Chlorella. A Beckman Model 864 infrared analyzer operated at its most sensitive range, 0-500 ppm, provided the analysis for  $\rm CO_2$ . Because the proper use of this equipment required that the test gas be passed through at approximately 100 cc/min, and because the flow rate in our experiments was only 10 cc/min, it was not possible to monitor the  $\rm CO_2$  in the effluent on a continuous basis. The air was collected, therefore, in a 100-cc syringe opened at a rate which maintained constant (atmospheric) pressure within the tube; the syringe was then taken from the

apparatus and the air expelled through a small drying column into the analyzer. The equipment used is represented in Figure The glass tube was 121.9 cm (4 feet) long and contained 85 Air bubbles were admitted through a glass ml of test water. tube, 2 mm ID, sealed into the throat of a ball joint and extending 2 mm above the opening in the joint. The ball joint was mated with the appropriate socket which was, in turn, fused to the bottom of the long glass tube. At the top of this tube was a plastic ell connected to a T-tube, the center leg of which extended into a reservoir of dilute acid. The connection from the other end of the T-tube to the glass syringe was made with a ball and socket joint. In taking the air sample, the plunger of the syringe was pulled open at the rate required to keep the meniscus in the dilute acid at its rest point. time required to fill the syringe was measured to give the air The four valves shown at the bottom of the diagram controlled the source and flow rate of air. Thermostated water was passed through the jacket surrounding the tube at a constant rate.

Prior to the start of an experiment the pressure regulators on each air cylinder were adjusted to 20 psi; a pressure of exactly 10 psi was maintained on the gauge shown in the figure by opening the valve leading to the proper air cylinder. rate through the glass tube was maintained by the needle valve at the lower right portion of the diagram, and air was allowed to escape at a constant rate through the valve at the top right for CO2 analysis. With this arrangement, there was no disturbance of the air/water equilibrium when the need arose for an analysis. To change from one CO2 concentration to another required only the closing of one valve and the opening of another, which took less than eight seconds. Experiments were of two types: in one, the water was first equilibrated (i.e., nearly so) with 0 ppm CO2 after which 346 ppm CO2 air was bubbled through and the other was the converse, with initial aeration with 346 ppm CO2 followed by 0 ppm CO2.

Some of the experiments included pH measurements which required that the connection at the top of the glass tube be modified to accommodate a Markson electrode which was connected with an Orion Model 801 meter.

The formula used to calculate the absorption (or release) rate of  ${\rm CO}_2$  is shown below:

$$cc CO_2/min = \Delta CO_2 (ppm) x air flow rate (cc/min)$$

$$10^6$$

The principal results of these equilibration studies are represented in the next four figures. Figure 12 is a plot of pH and the volume of  $CO_2$  released per minute from a 25% seawater solution (salinity = 8.24 ppt) which had been initially at equilibrium with 346 ppm  $CO_2$  but then aerated with 0 ppm

 ${\rm CO}_2$ . The starting pH was 8.01 and in this case the final was 9.00, but the upper pH limit is a function of aeration time and not nearly so well defined as that when equilibrium with air containing  ${\rm CO}_2$  is established. Figure 13 represents data from the converse experiment in which 25% seawater, initially almost at equilibrium with 0 ppm  ${\rm CO}_2$ , was aerated with 346 ppm  ${\rm CO}_2$ ;  ${\rm CO}_2$  uptake was complete in about five hours and the pH ended up exactly at the starting point for the previous experiment.

To demonstrate conveniently the relative rates of  $\rm CO_2$  absorption as a function of salinity requires that an arbitrarily selected rate of absorption be used as the starting point of the plot, otherwise the crisscrossing of lines with varied slopes becomes confusing. In Figure 14, the zero time point corresponds to an uptake rate of 2.56 x  $10^{-3}$  cc/min; beyond that point, only 200 minutes were required for equilibrium to be established with 25% seawater but more than 800 minutes were required with the 100% seawater.

With the technique described here it was easy to measure the uptake or release per minute of  $\rm CO_2$  amounting to only 0.0026% of the total dissolved  $\rm CO_2$ . These figures refer to undiluted seawater and an air flow rate of 10 cc/min.

Figures 12 and 13 demonstrate the relationships between pH and release, or absorption, of CO2 by a 25% seawater solution which has either been saturated or stripped of CO2. Time zero in these plots marks the changeover from one air cylinder to another. An appreciation of the relative sluggishness of the pH response compared to that of gas exchange is demonstrated by a different use of the data used in Figure 13. Figure 15 contains data on pH and CO2 prior to, and after, the change from one air stream to another and the plot of CO2/minute includes both negative and positive values. Prior to time zero there was a very slow release of CO2 though the stripping operation had been proceeding overnight, and during this time the pH rose at a rate of about .001 unit in 5 minutes. With the change at time zero, from 0 ppm to 346 ppm CO2, the pH continued to rise for several minutes before starting to drop slowly. Concurrent with the slow pH change was an enormous change in the composition of the air exiting from the system. At this point the minimum pH change corresponded with a maximum change in CO2 absorption.

The complexity of the equilibration of  ${\rm CO}_2$  with waters of various salinities is demonstrated in the figures shown. Rates of change of  ${\rm CO}_2$  uptake or release depend upon both pH and salinity. To summarize the research on limitations of the gas exchange approach, with  ${\rm CO}_2$  in waters of various salinities, the salinity should probably be below 13 parts per thousand otherwise the gas exchange rate is too slow for meaningful measurements.

### IV. DISCUSSION

The thrust of this report is to emphasize the speed with which changes take place in the  $O_2$  or  $CO_2$  content of air passing through cultures of microorganisms, compared to changes in biomass. Three factors prompted an awareness of this phenomenon in the research at NRL:

- a. The Chlorella used in the algal studies had an extraordinarily high O<sub>2</sub> production rate. The six-lamp unit could produce over 700 cc O<sub>2</sub>/liter/hr and another unit featuring an illuminated area only 1 cm deep produced more than double that amount.
- b. High air flow rates could be used to advantage in such cultures which, in turn, resulted in a fast equilibration between the air and the liquid.
- c. The Chlorella cultures had a low salinity which also was responsible for fast air/liquid equilibration.

In this report the emphasis has been on the rapidity with which changes in 02 production of an algal culture can be detected following the imposition of physical or chemical changes on the system. Pressure, CO2 input rate, light intensity, and the presence of a toxic alloy were shown to induce practically immediate responses in terms of the O2 production rate, whereas nutrient changes (either by the addition of extra urea, or by the substitution of a particularly concentrated culture medium) resulted in more gradual changes in O2 production. Neither packed cell volume nor cell number is a sensitive measure of the productivity of an algal culture, largely because the  $O_2$  production rate is a function of illuminated area of the culture once enough cells are present to make the culture light-limited. Therefore, biomass changes of an algal culture are often unsatisfactory criteria of its performance; in contrast, the changes in O2 production rate can be remarkably fast and easily detected.

The instances cited have concerned either  $0_2$  or  $C0_2$  as components of an air stream but the concept is not limited to these two gases. The same consideration would apply to any gas for which a sensor is available.

### Reproducibility

The most compelling argument for the reliability of an algal culture as a producer of  $O_2$  rests on the results obtained with the six-lamp unit over a five-day period, when the total variation in results could be accounted for by misreading the  $O_2$  analyzer by only 0.01%. Under controlled conditions of light intensity,  $CO_2$  aeration rates, etc., algal cultures will behave

predictably. The responsibility of the researcher is to ensure that the effluent air being analyzed is representative of air at equilibrium with the system. The main concern here is air flow rate; if, for some reason, the air flow rate were reduced momentarily the result would be a change in the air composition. It is necessary, therefore, to keep the flow rate constant and to allow enough time for the effluent to sweep out the volume contained in the drying column and the analyzer before a reading is taken.

The reproducibility of gas exchange studies was demonstrated also in the studies of equilibration of  $\text{CO}_2$  with 25% seawater. Three experiments were performed in which the seawater was equilibrated with 346 ppm  $\text{CO}_2$  after an initial aeration with 0 ppm  $\text{CO}_2$ . The initial pH's were different but the results obtained, relating  $\text{CO}_2$  uptake vs. time, gave super-imposable plots when adjusted to a fixed uptake value.

Temperature can be an important factor. In the studies concerning equilibration with dilute seawater, it was necessary to determine the point when equilibrium had been reached, i.e., when the effluent air had the same  $\mathrm{CO}_2$  content as the input. Instances in which the equilibrium was obviously delayed could usually be traced to minor changes in temperature. An elaborate system of temperature control, which limited the variation to less than  $0.06\,^{\circ}\mathrm{C}$  during the course of an experiment, solved the problem. The problem was less acute with the large algal culture unit, perhaps because at its temperature (39 $^{\circ}\mathrm{C}$ ) the rate of change of  $\mathrm{CO}_2$  solubility per degree centigrade is less than half that at  $22\,^{\circ}\mathrm{C}$ .

Bubble size can be an important factor. It was not a focal point in these studies but an insight into its importance was obtained when one bubbler was substituted temporarily for another in the  ${\rm CO}_2$  equilibration studies. The difference in the diameters of the two tubes was not apparent to the eye but the gas exchange characteristics were greatly different.

### Additional Factors to be Considered

There may be a misconception that the  $O_2$  production of an algal culture cannot be constant and that it must increase as growth proceeds. That happens to be the case with extremely dilute cultures when all the cells are subjected to the same light intensity at all times. This was not the situation with the rather dense cultures (greater than 1% packed cell volume) used in the Chlorella studies where only a thin layer of cells closest to the light received the full intensity; as mentioned earlier, in such "light limiting" conditions the productivity of the culture is a function of the area of illumination and the  $O_2$  production is constant over quite a wide range of cell concentrations.

To this point, mention has been made of  $\mathrm{CO}_2$  and  $\mathrm{O}_2$  measurements as single entities but in many instances the combination of the two can provide additional information. The assimilatory quotient for algae  $(\mathrm{O}_2/\mathrm{CO}_2)$  varies according to the nitrogen source being utilized by the algae. This is well known and will not be explored further at this point. What, perhaps, is not so well known is the variation of the assimilatory quotient with the condition of the organism. The Chlorella cultures, with urea as the nitrogen source, had an AQ of 1.06 when all conditions were favorable for growth. But a culture which had been refrigerated over a weekend would have an AQ of 0.9 or even less for at least several hours in the mass culture apparatus. It is useful, therefore, to know what the expected AQ is in order to make a judgment about the well being of the culture at any time.

The better known respiratory quotient (CO2 produced/O2 absorbed) has been a useful diagnostic in medical research on humans and animals, though there are instances in which its particular relevance have been overlooked. A case in point [work not performed at NRL, obviously] concerns the effect of certain aflatoxins occurring in the feed given to chickens; for a period of several weeks' exposure to these compounds the chickens have no visible defects, but with the routine handling associated with the killing of the chickens and removal of their feathers, the blood vessels hemorrhage, resulting in a considerably lower sale price of the meat. Clinical investigations have shown that an effect of the aflatoxins is to decrease the ability of the chickens to absorb fat from their diet, and fats in the feed pass through the animal into the feces without digestion. The difference between the respiratory quotient of a healthy chicken, and one exposed to aflatoxins, should be detectable and could serve as a warning that an abnormal condition existed.

### Alloy Toxicity

The most intriguing research included in this report concerns the inhibition of O<sub>2</sub> production by Chlorella when exposed to 3.8 mm diameter rods made of certain alloys. Dense cultures (approximately 1.5% packed cell volume) were used in this study and two questions come to mind when contemplating such remarkable results: 1) why is the O<sub>2</sub> production changed so abruptly, and to a new constant level, rather than a slow decline which one might expect with the imposition of a dilute toxic agent?; 2) how can such a dense culture be affected by what must be minor concentrations of extractable metallic species?

The answers to these questions are not known. In that particular study, an attempt was made to duplicate the decline in  $O_2$  production by introducing varying concentrations of metal ions of the alloy components ( $Cr^{3+}$ ,  $Cr^{6+}$ , etc.) but

the results were not definitive. Also, the concentrations required to produce such inhibitions were too high to be regarded as realistic values. The results obtained with one rod would support the notion that leakage of some particularly toxic component from a grain boundary was the source of the inhibition. Perhaps an exotic species vastly different from the single ion components tested could be responsible for the inhibitions noted.

In all deliberations on this subject, the focus has been on toxicity. It may be more accurate to regard changes in  $O_2$  productivity as the result of a change in synchrony of the cells. Sorokin (1963) showed (Figure 16) the changing rates of photosynthetic gas exchange for ten-minute intervals in cells that were synchronized. The new (0-hr) cells showed a progressive rate of gas exchange while the older (8-hr) cells showed a decline. Therefore, a drastic change in synchrony, in a population which is totally diverse in terms of cell age, could result in a change of  $O_2$  production rate. Once again, however, is the problem that an explanation be given for the release of sufficient material from the metal alloy in question to cause a change in synchrony of the cells.

For the purpose of this discussion it is not necessary to resolve the dilemma although it would certainly be desirable. The essential point is that an algal culture can respond to a specific metal alloy specimen in a very short time which presents an opportunity for exploitation. A likely possibility concerns the study of marine corrosion, and the connection between corrosion and algal susceptibility to toxicants is based on the following line of thought: a) inhibition of 02 production by an alloy is a surface phenomenon; b) leaching of constitutents from a grain boundary in the surface is a plausible explanation for the effect of an alloy on an algal culture; c) resistance to corrosion by an alloy may be reduced by the loss of the same constitutents. If the premise is valid, then an initial screening test of alloy specimens for toxicity (particularly the stainless steels) might be correlated with corrosion susceptibility in a field test. This is admittedly a simplistic explanation for what must be a difficult field of study, but metallurgists seeking an explanation for the occasional, unusual, corrosion problem might consider this possibility.

Convincing arguments for the greater use of gas exchange studies could be generated by researchers in diverse fields who are aware of the existing problems in those fields. The instances of applications cited here concern algae for the most part, but also include animal experimentation and metallurgical research. Additional work being done at NRL, but not included in this report, concerns the  $\rm CO_2$  production rate of baker's yeast as the central point of an assay for the detection of certain toxins. Preliminary results are extremely promising in that, in certain instances, the effect of only 2 micrograms

of toxin is apparent in only 12 minutes.\* All of these methods have in common the need for control over air flow rate and temperature and a  $\rm CO_2$  or  $\rm O_2$  analyzer. The costs for equipment are modest compared to so much of the experimentation which is found in most research laboratories. The essence of the gas exchange approach is that each measurement is a rate measurement -- those who have an appreciation of this fact should be interested in further studies.

### V. REFERENCES

- 1. J. D. Bultman and P. J. Hannan, "A Continuous Gas-Mixing Apparatus," J. Chem. Ed., 44, 424, (1967).
- 2. D. L. Dyer and D. E. Richardson, "Materials of Construction in Algal Culture," Appl. Microbiol., 10, 129 (1962).
- 3. P. J. Hannan, "Effect of Pressure on Oxygen Production by Algae," Dev. Ind. Microbiol., 6, 229 (1964).
- 4. P. J. Hannan and C. Patouillet, "Gas Exchange with Mass Cultures of Algae. I. Effects of Light Intensity and Rate of Carbon Dioxide Input on Oxygen Production," Appl. Microbiol., 11, 446 (1963).
- 5. P. J. Hannan and C. Patouillet, "Gas Exchange with Mass Cultures of Algae. II. Reliability of A Photosynthetic Gas Exchanger," Appl. Microbiol., 11, 450 (1963).
- 6. P. J. Hannan and C. Patouillet, "Toxicity of Metals to Chlorella--A Surface Phenomenon," Dev. Ind. Microbiol., 8, 313 (1967).
- 7. P. J. Hannan and C. Patouillet, "An Algal Toxicity Test and Evaluation of Adsorption Effect," J. Wat. Poll. Cont. Fed., 51, 834 (1979).
- 8. P. J. Hannan, R. L. Shuler and C. Patouillet, "A Study of the Feasibility of Oxygen Production by Algae for Nuclear Submarines," NRL Report 5954 (1963).
- 9. J. Lyman and R. H. Fleming, "Composition of Seawater," J. Mar. Res., 3, 134 (1940).

<sup>\*</sup>Funding for this project has been obtained from the U.S. Army Medical Research and Development Command, Fort Detrick, MD.

- 11. C. Sorokin and J. Myers, "A High Temperature Strain of Chlorella," Science, 117, 330 (1953).
- 10. C. Sorokin, "On the Variability in the Activity of Photosynethic Mechanisms" from "Photosynthesis Mechanism in Green Plants," Publication 1145, Natl. Acad. of Sciences (1963).

Table 1 - Burk's medium

Major Constituents	Amount (grams/liter)
MgSO₄ 7 H₂O	5.0
KH <sub>2</sub> PO <sub>4</sub>	2.5
NaCl	2.0
Urea	0.4

Micronutrients	Conc. Source Compd. (mg/liter)	Conc. Trace Element (ppm)	M.E.Q. Element (liter)
Fe - EDTA	4	l Fe	5.36 × 10-1
CaCl <sub>2</sub>	22	8 Ca	.8
H <sub>3</sub> BO <sub>3</sub>	5.7	1 B	.6
MnCl <sub>2</sub> 4 H <sub>2</sub> O	3.6	l Mn	3.64 × 10-2
ZnSO.	0.44	0.18 Zn	5.5 × 10 <sup>-2</sup>
CuSO₄ 5 H₂O	0.158	0.04 Cu	1.25 × 10-2
(NH <sub>4</sub> ) <sub>6</sub> Mo <sub>7</sub> O <sub>24</sub> 5 H <sub>2</sub> O	0.035	0.019 Mo	1.19 × 10-3
NaVO <sub>3</sub>	. 2	0.84 V	8.24 × 10 <sup>-2</sup>

Table 2 - Summary of week-long performance of six-lamp unit

Light Intensity: 23,000 Foot-Candles Electrical Power: 7040 Watts

Dilution Rate: 15.6% per hour Gas Flow Rate: 4000 cc/min, containing

1.9% CO.

		Oxygen F	Oxygen Production	
Бау	Suspension Density Percent Packed Cell Volume	Volume (cc/hr)	Standard Deviation (cc/hr)	Dilution Rate (ml/hr for 24 hr)
Monday	1.27 - 1.37	3432	145	Not measured
Tuesday	1.44 - 1.50	3565	30	964
Wednesday	1.43 - 1.47	\$606	29	696
Thursday	1.41 - 1.44	3584	25	996
Friday	1.41 - 1.43	3638	24	950

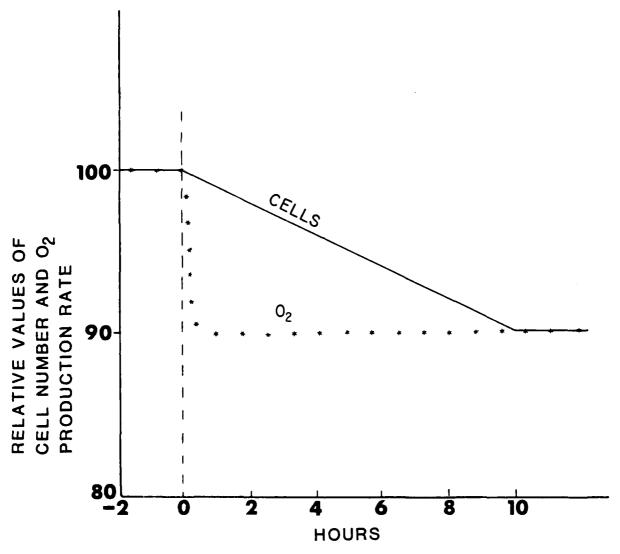


Fig. 1 — This is a theoretical plot of the cell number and the  $O_2$  production rate, vs time, of an algal culture being diluted with fresh medium at 10% hr<sup>-1</sup>. At time zero the growth rate was reduced 10% from the original.

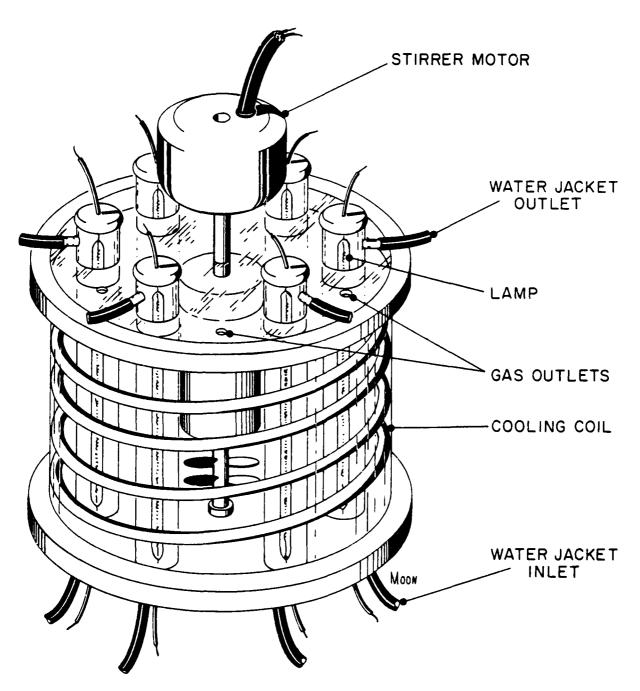


Fig. 2 — Schematic representation of a mass culture unit containing six Quartzline high intensity lamps

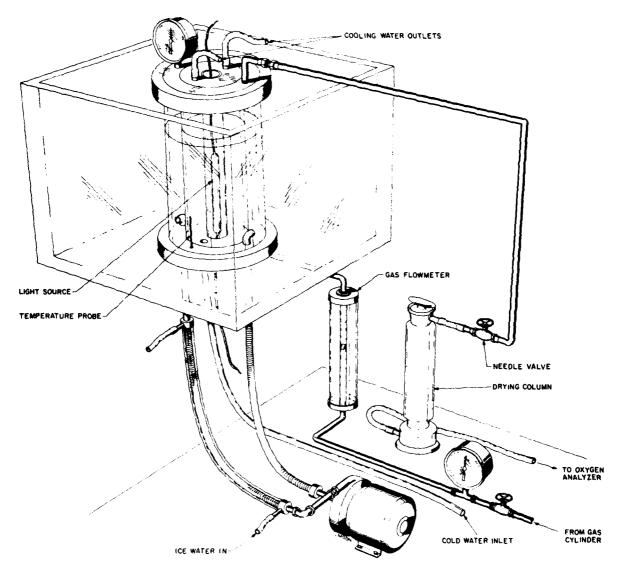
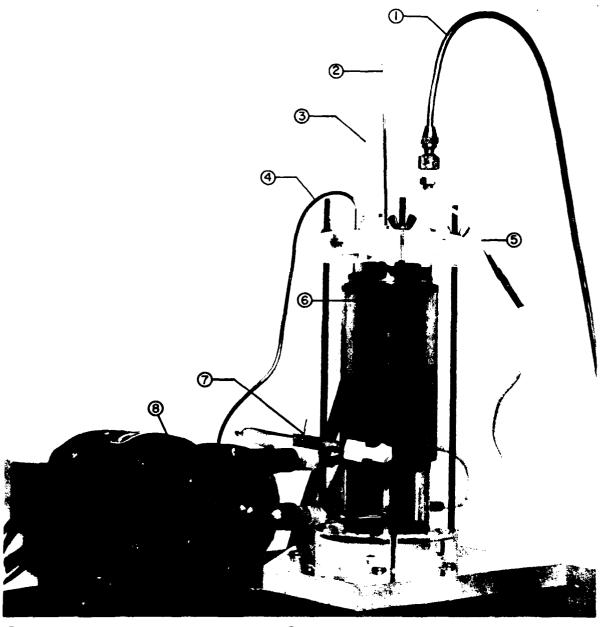


Fig. 3 — Diagram of a culture apparatus, made from glass pipe and end plates of Plexiglas, which was used for the study of pressure effects on *Chlorella*. An inner annular space contained water to dissipate the heat released by the Quartzline lamp extending through the unit. The algal culture, in the outer annulus, was circulated by a centrifugal pump underneath this unit.



- ( AIR INLET TUBE
- @ METAL SAMPLE ROD
- 3 COOLING JACKET OUTLET
- 4 TEMPERATURE SENSING PROBE
- 5 POWER CABLE FOR LIGHT SOURCE
- 6 OUTER JACKET OF ALGAE CULTURE UNIT
- 7 COOLING JACKET INLET
- 8 ALGAE CIRCULATING PUMP

Fig. 4 — Algal culture unit used in toxicity studies. A light source extended vertically through the unit, surrounded by annular spaces containing cooling water and the algal culture; circulation of the culture was provided by the centrifugal pump. Additional cooling of the culture was provided by cold water, circulating through an outer jacket, on demand from a thermoregulator probe. A 1/8" diameter rod is shown extended above the unit; it is held in place by an O-ring seal.

# EFFECT OF LIGHT INTENSITY ON O2 PRODUCTION

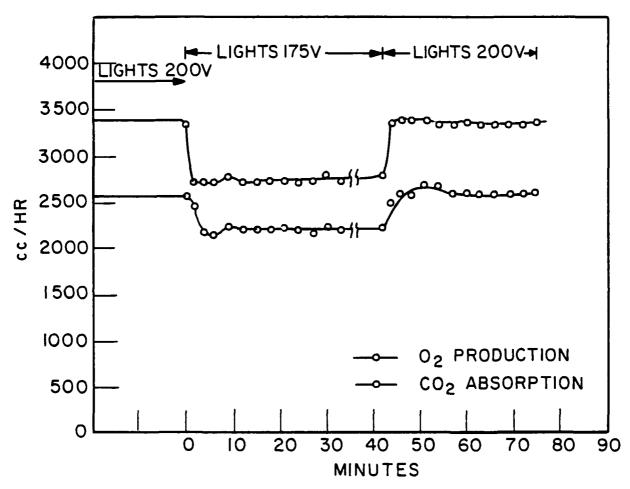


Fig. 5 — A plot of the O<sub>2</sub> production and CO<sub>2</sub> absorption rates of mass culture of *Chlorella* in response to light intensity changes.

## EFFECT OF CO2 SUPPLY RATE ON GAS EXCHANGE

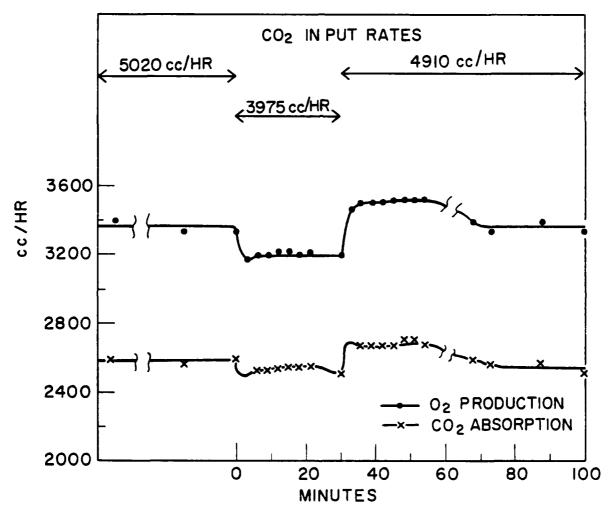


Fig. 6 – A plot showing the relationship between O<sub>2</sub> production, and CO<sub>2</sub> uptake, of a mass culture algae with changes in the CO<sub>2</sub> input rate.

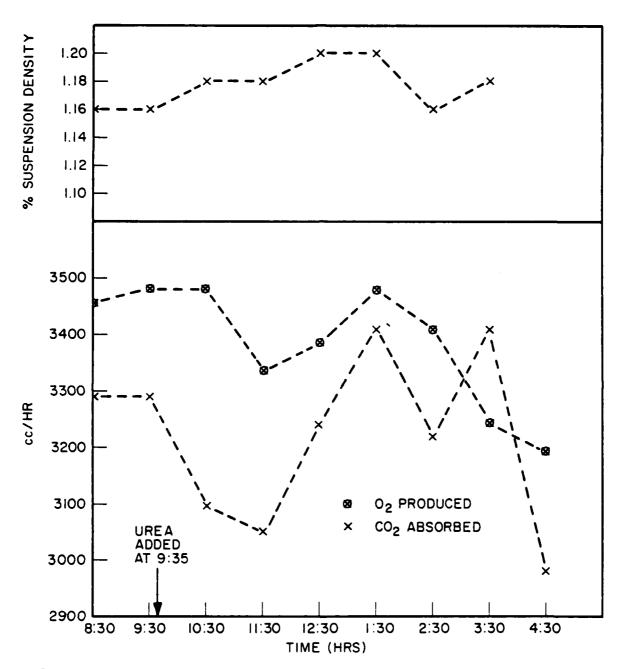


Fig. 7 — The effect of doubling the urea concentration of *Chlorella* which was being diluted at a constant rate of 15% hr<sup>-1</sup> with regular culture medium. Note the slight changes in suspension density in comparison with the large excursions in  $CO_2$  absorption and  $O_2$  production.

# ADAPTATION OF CELLS TO ENRICHED MEDIUM

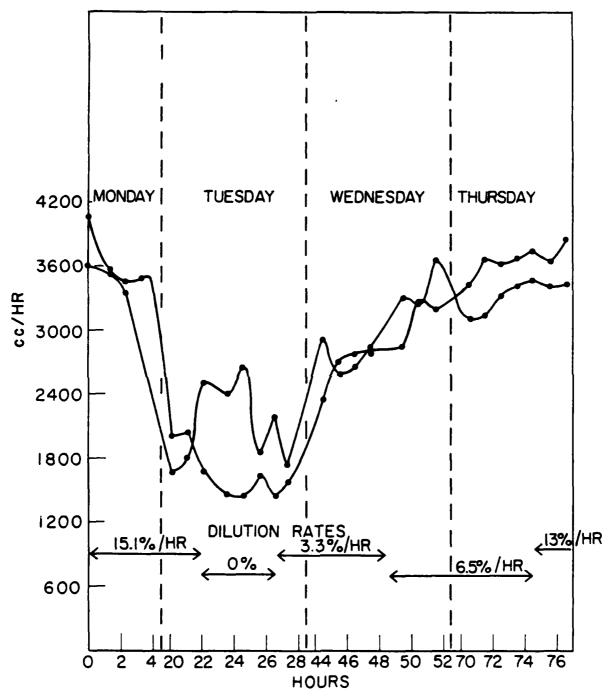


Fig. 8 — Variations in O<sub>2</sub> production and CO<sub>2</sub> uptake of mass culture of *Chlorella* following the substitution of a highly concentrated medium for the one normally used.

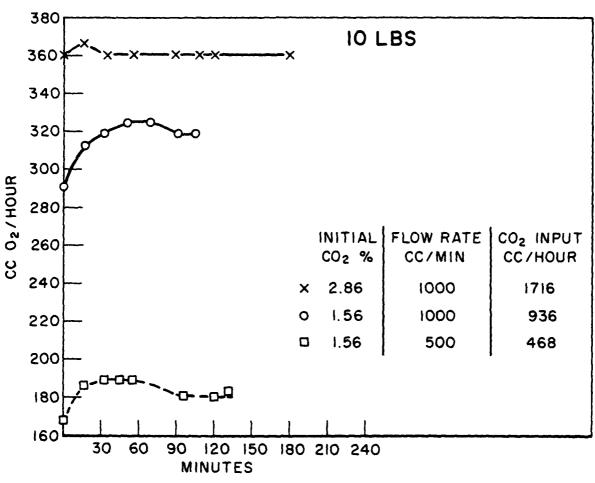


Fig. 9 — Plots showing the effects of various pressures, at three input rates, on  $O_2$  production by *Chlorella*.

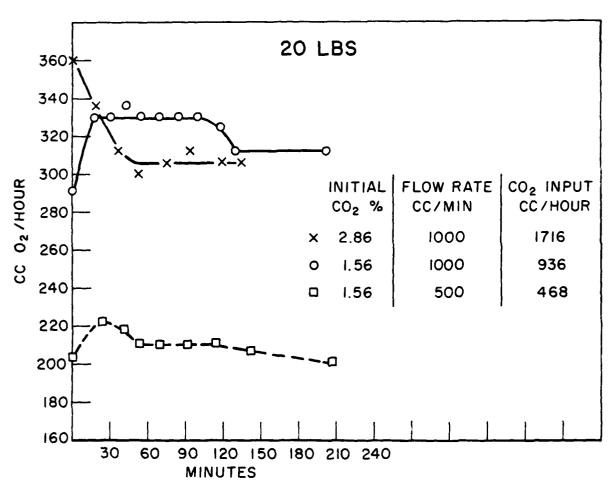


Fig. 9 (Cont'd) — Plots showing the effects of various pressures, at three input rates, on  $O_2$  production by *Chlorella*.

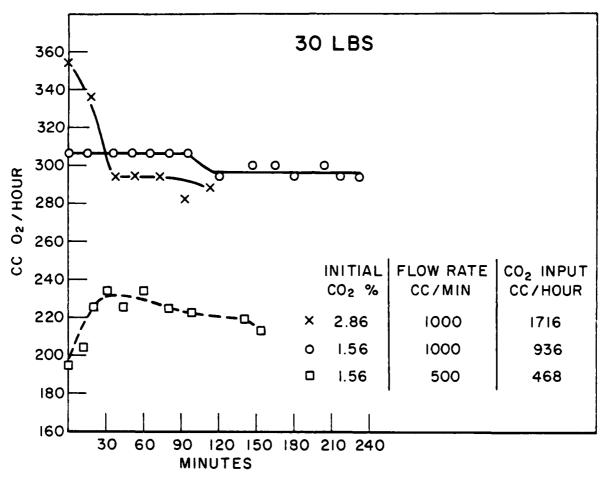


Fig. 9 (Cont'd) — Plots showing the effects of various pressures, at three input rates, on O<sub>2</sub> production by *Chlorella*.

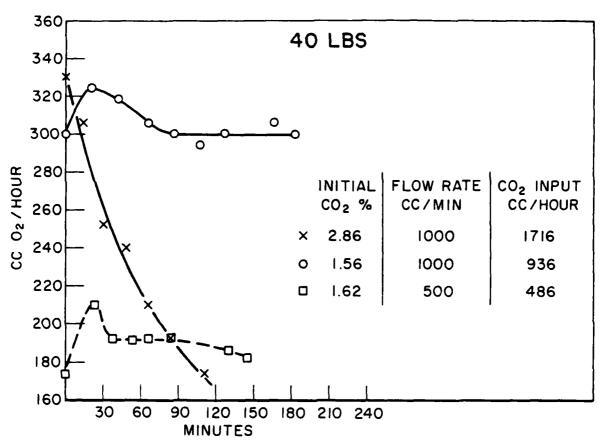


Fig. 9 (Cont'd) — Plots showing the effects of various pressures, at three input rates, on O<sub>2</sub> production by *Chlorella*.

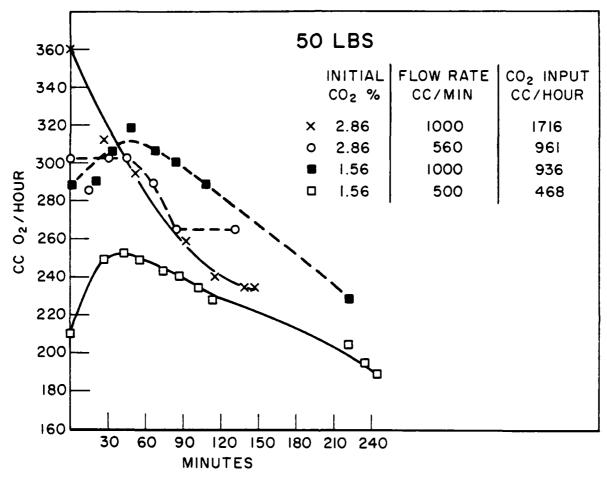


Fig. 9 (Cont'd) — Plots showing the effects of various pressures, at three input rates, on O<sub>2</sub> production by *Chlorella*.

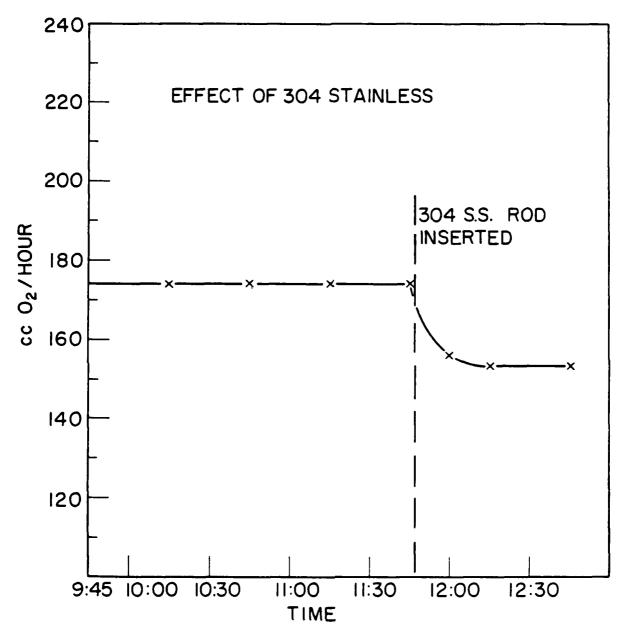


Fig. 10 — Oxygen production of an algal culture before and after the insertion of a 1/8" diameter rod of #304 stainless steel into the culture. In order to provide an expanded scale, the plot on the vertical axis is not extended to zero.

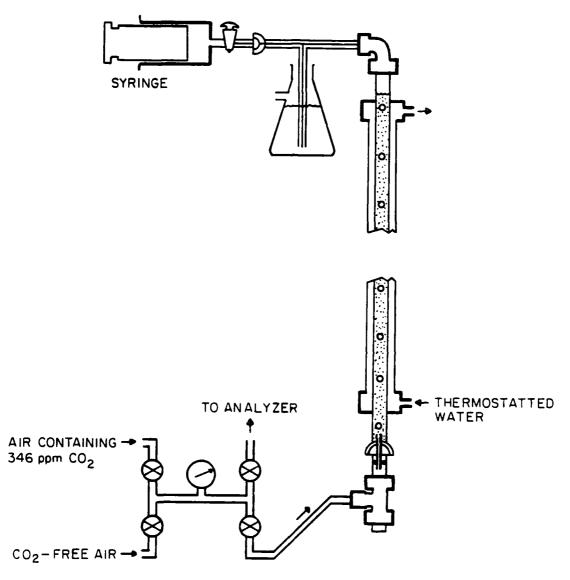


Fig. 11 — Schematic diagram of an apparatus used to study equilibrium rates of  $\rm CO_2/air$  mixtures with seawater of various salinities.

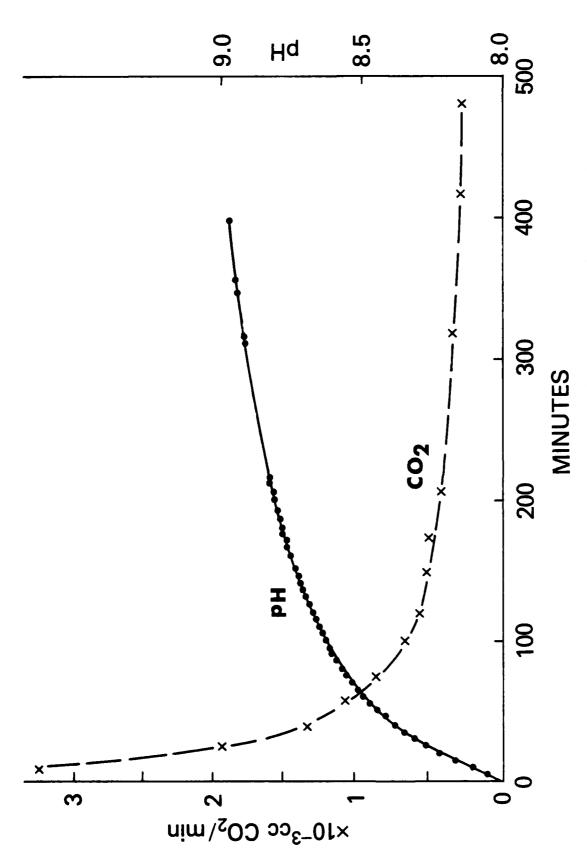


Fig. 12 — A plot of CO<sub>2</sub> release, and rise in pH, of a 25% seawater solution from the time of introduction of 0 ppm CO<sub>2</sub>. The solution had been in equilibrium with 346 ppm CO<sub>2</sub>.

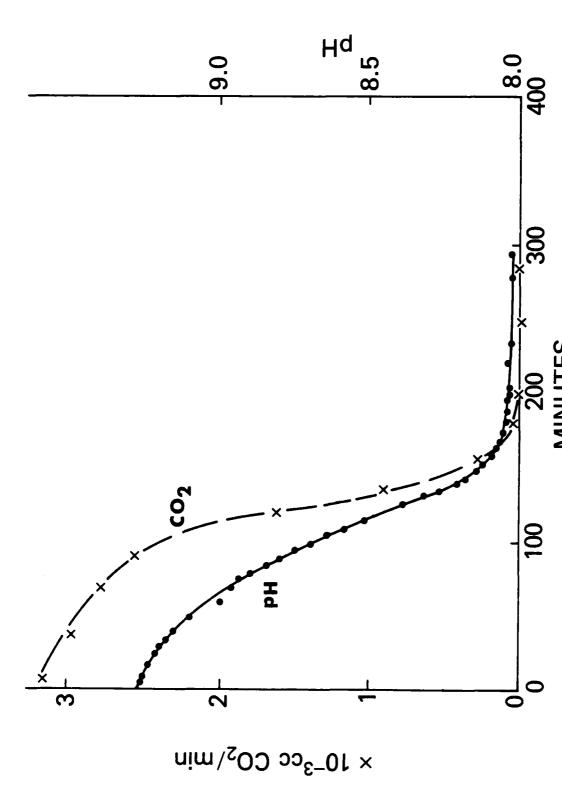
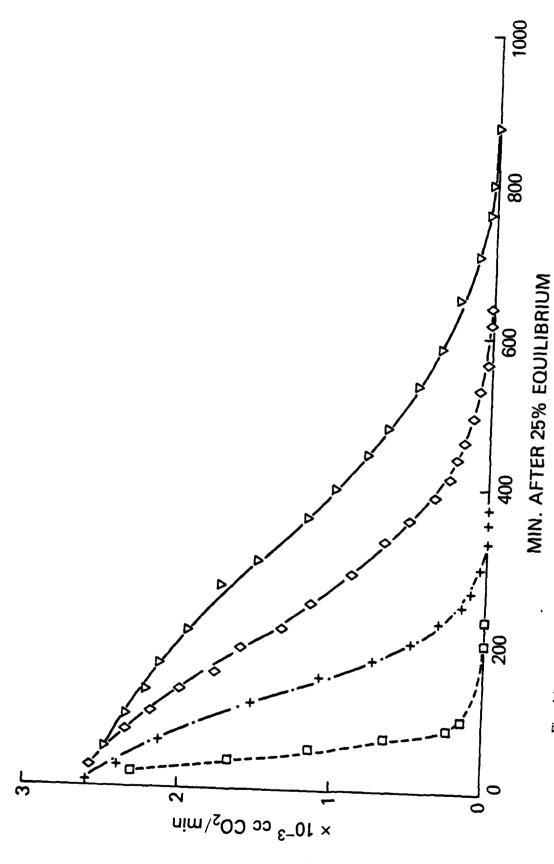


Fig. 13 — A plot of the CO<sub>2</sub> uptake, and decline in pH, of a 25% seawater solution from the time of introduction of air containing 346 ppm  $CO_2$ . The solution had been aerated overnight with  $CO_2$ -free air.



いいから、これないないない。

Fig. 14 — This plot illustrates the rates of CO<sub>2</sub> uptake, with time, by various seawater solutions exposed to 346 ppm CO<sub>2</sub> after having been initially almost at equilibrium with 0 ppm CO<sub>2</sub>. The air flow rate was 10 cc/minute and the origin of the plots was taken to be an arbitrary rate of 2.56  $\times$  10<sup>-3</sup>

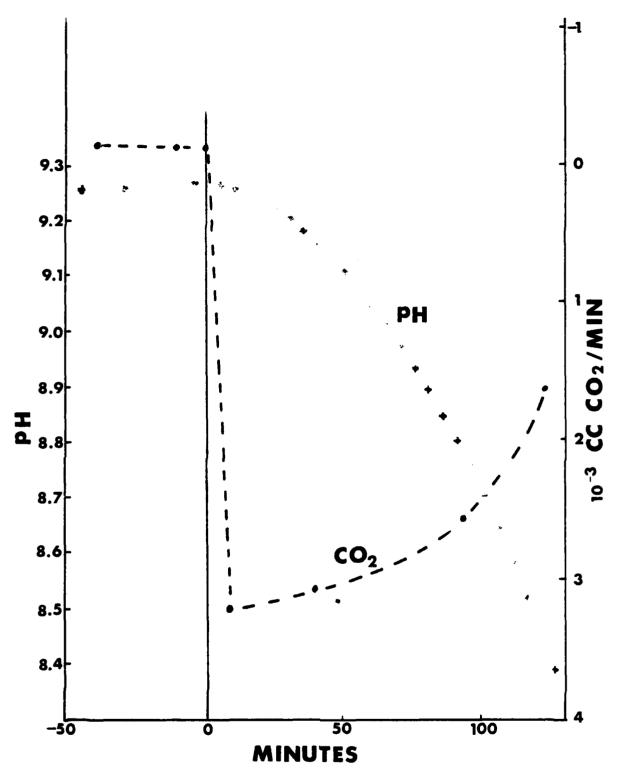


Fig. 15 — A plot showing the relationship between pH and  $CO_2$ , before and after the substitution of air containing 346 ppm  $CO_2$  for that containing 0 ppm  $CO_2$ . The liquid was 25% seawater. Note the gradual change in pH vs the drastic initial absorption of  $CO_2$ .

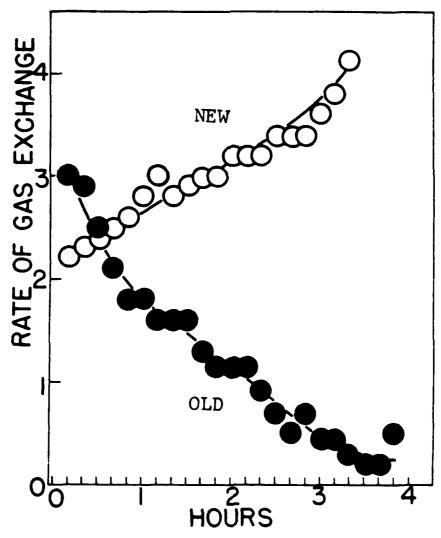


Fig. 16 — Rates of photosynthetic gas exchange (in relative units) for 10-minute intervals in synchronized 0- and 8-hour cells of *Chlorella*. Taken from Publication 1145, National Academy of Sciences, "On the Variability in the Activity of Photosynthetic Mechanisms." Printed with permission of the Academy.

Appropriate transcent because the property in the second sections.

# EMED

5-86 DT [